

**Figure S1.** Multigating strategy for the analysis of surface antigens and intracellular cytokines of mononuclear cells. Lymphocytes (R1) were identified based on gating of forward scatter area (FSC-A) and side scatter area (SSC-A). And then forward scatter height (FSC-H) and FSC-A gating was used to obtain single lymphocytes (R2). Total live lymphocytes (R3) were then identified based on the exclusion of a Live/Dead dye (FVS780). Positive staining for CD3 was then used to distinguish live T lymphocytes (R4). Subsequently, representative plots showed percentage of CD4+ T lymphocytes (R5), CD8+ T lymphocytes (R6), CXCR3+ T lymphocytes (R7), Ki67+ T lymphocytes (R8), IFN-+ T lymphocytes (R9), IFN-+CXCR3+ T lymphocytes (R10), IFN-+CD8+ T lymphocytes (R11), IFN-+CD4+ T lymphocytes (R12), and IFN-+Ki67+ T lymphocytes (R13) within all the live T lymphocytes.



**Figure S2.** Body weight changes of mice (A and B) and guinea pigs (C) in this study. Values greater than zero represent weight gain, and values less than zero represent weight loss. Group data showing that there is no significant difference between different groups.



**Figure S3.** **(A)** Group data showing the ratio of IFN-+CD8+ T lymphocytes to IFN-+CD4+ T lymphocytes in spleen T lymphocytes. **(B)** Group data showing the ratio of IFN-+CCXR3+ T lymphocytes to IFN-+ T lymphocytes in spleen T lymphocytes. Mice in different groups were treated as the same as that in Figure 2. n = 6-8 per group. Data are shown as mean ± SD. ＊, *P* < 0.05, compared with the “Control”. ＃, *P* < 0.05, compared with the “IFN- model”.



**Figure S4.** Effects of IFN- and different doses of CXCR3 inhibitor on blood lymphocytes. **(A1-A5**) Representative plots showing the proportion of IFN-+ T lymphocytes in blood T lymphocytes from the group of “Control” (A1), “IFN- model” (A2), “low-dose CXCR3 inhibitor” (A3), “medium-dose CXCR3 inhibitor” (A4), and “high-dose CXCR3 inhibitor” (A5). **(A6)** Summarized data showing the proportion of IFN-+ T lymphocytes in blood T lymphocytes. **(B)** Summarized data showing the proportion of CXCR3+ T lymphocytes in blood T lymphocytes. **(C)** Group data showing the ratio of IFN-+CCXR3+ T lymphocytes to IFN-+ T lymphocytes in blood T lymphocytes. **(D)** Group data showing the ratio of IFN-+CD8+ T lymphocytes to IFN-+CD4+ T lymphocytes in spleen T lymphocytes. **(E)** Group data showing the proportion of Ki67+ T lymphocytes in spleen T lymphocytes. Mice in different groups were treated as the same as that in Figure 2. n = 6-8 per group. Data are shown as mean ± SD. ＊, *P* < 0.05, compared with the “Control”. ＃, *P* < 0.05, compared with the “IFN- model”.



**Figure S5.** **(A)** Group data showing the ratio of IFN-+CD8+ T lymphocytes to IFN-+CD4+ T lymphocytes in lung-resident T lymphocytes. **(B)** Group data showing the ratio of IFN-+CCXR3+ T lymphocytes to IFN-+ T lymphocytes in lung-resident T lymphocytes. **(C)** Group data showing the proportion of Ki67+ T lymphocytes in lung-resident T lymphocytes. Mice in different groups were treated as the same as that in Figure 2. n = 6-8 per group. Data are shown as mean ± SD. ＊, *P* < 0.05, compared with the “Control”. ＃, *P* < 0.05, compared with the “IFN- model”.



**Figure S6.** **(A**) Effects of IP-10 on the proliferation of blood mononuclear cells were assessed by CCK-8 assay. **(B**) Effects of IP-10 on the proliferation of blood lymphocytes were counted by flow cytometry. **(C**) Effects of IP-10 on the proliferation of lung-resident mononuclear cells were assessed by CCK-8 assay. **(G**) Effects of IP-10 on the proliferation of lung-resident lymphocytes were counted by flow cytometry. n = 6 per group. Data are shown as mean ± SD.